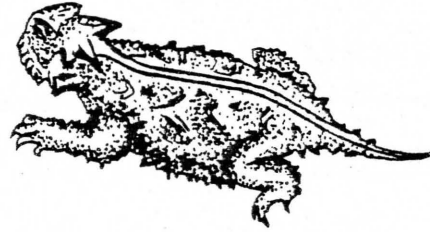




KANSAS
HERPETOLOGICAL
SOCIETY



NEWSLETTER

NUMBER 35

FEBRUARY 1980

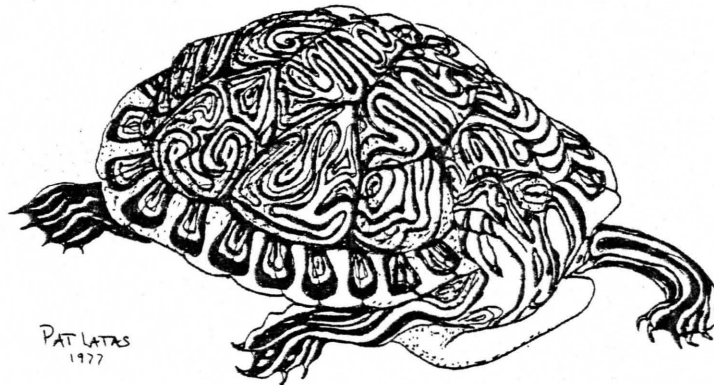
MARCH KHS MEETING TO BE HELD IN WICHITA

The next meeting of the Kansas Herpetological Society will be held on Saturday, March 22, at the beautiful Sedgwick County Zoo in Wichita. The program for the day will consist of the following:

- | | |
|-----------------|--------------------------------------------------------------------------------------------------------------|
| 9:30 - 10:00 AM | Coffee and doughnuts in the Public Education Room |
| 10:00 - Noon | Lecture: "Psex and the Psingle Pseudemys." by Fredric Frye, D.V.M. |
| Noon - 12:15 PM | Lecture: "Turtle farming in Grand Cayman" by Barbara Paschke, KU Office of Institution Research and Planning |
| 12:15 - Noon | LUNCH |
| 1:00 - 2:00 PM | A "Behind the Scene" guided tour of the Herpetarium with Eddie Stegall |

An Executive Council meeting will be held during the noon hour. We are extremely lucky to have Dr. Frye, a leading authority on the proper care of captive reptiles and their diseases, speak to us at this meeting. He is the author of a very informative book, containing numerous color plates, entitled: Husbandry, Medicine and Surgery in Captive Reptiles. 1973. VM Pub., Inc., Bonner Springs, KS. In addition, Barbara Paschke will share her experiences with us concerning turtle farming in the Caribbean. This is also an opportunity to visit the modern Sedgwick County Zoo. So bring your family and friends. In addition to the beautiful exhibits, you will have the unique opportunity to see the zoo's herpetarium as the zoo staff does. During my last tour, I was able to see an extensive project that was undertaken to breed poison-arrow frogs (Dendrobatidae).

Be sure to come to the March meeting, and, bring a friend.



FIRST KHS MEETING OF THE 80'S HELD AT EMPORIA

The threat of icy roads kept many members away from the first meeting of the year, held at Emporia State University. The afternoon began with conversation around the pot of hot coffee which was generously provided by Dr. Robert Clarke, of the host institution.

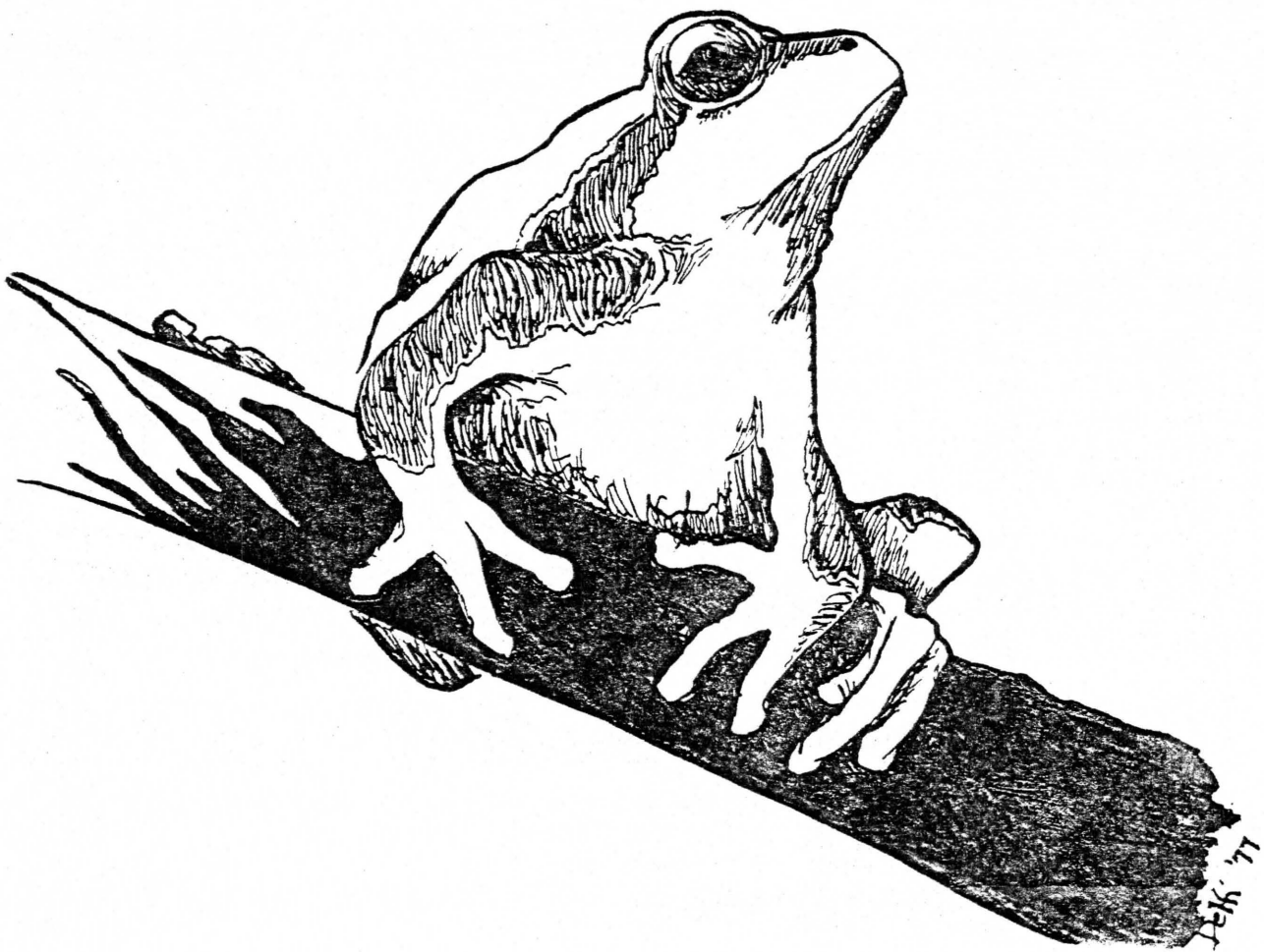
The meeting began with the presentation of a videotape prepared by Kansas Fish and Game, in which Joseph T. Collins discussed the beneficial nature of snakes in our environment. Various common snakes of Kansas were shown, so that the general public may learn to recognize them in the field. Special emphasis was placed upon knowing the poisonous snakes that occur in the state. The vividly colored red milk snake (Lampropeltis triangulum sypila) captured our attention, while Joseph Collins explained that because it superficially resembles coral snakes (Mirurus spp., Micruroides sp.) which are poisonous, it is often killed. Some of the other snakes that were shown include: the bullsnake (Pituophis melanoleucus sayi), the black rat snake (Elaphe obsoleta obsoleta), the timber rattler (Crotalus horridus), and the prairie rattler (C. v. viridis). It was a good presentation that was especially enjoyable since it was in color.

Next, Kelly Irwin gave a slide presentation of the beautiful islands that comprise American Samoa. The four islands, Ofu, Olosega, Tau, and Tutuila, are covered with lush tropical vegetation, and, are bordered by numerous spectacular coral reefs and rock formations which occur along their coastlines. The aggregated skeletons of the innumerable anthozoans which have died many years ago often take the shape of large, flat plates. These plates were used by the natives to construct an ancient walkway, 1000 to 1500 years old, along the tropical forest floor. Flying foxes were commonly seen eating the ripening papayas. A total of six species of skinks (family Scincidae) were seen on the islands, including Emoia nigra, a large, black skink found both in the trees as well as on the forest floor. While the adults are jet-black, the juveniles possess rows of orange dorsolateral spots, making them particularly striking. A terrestrial skink (Emoia lawesii) which often reaches a total length of eight inches, was seen on or beneath the large, coral plates of the ancient walkway. Another skink (E. samoense) which was predominantly green in color, occurred in the forest vegetation. Moving to the edge of the forest, Kelly encountered yet another skink species (E. cyanura) living on low shrubs near the ocean. In addition to the previously mentioned members of the Scincidae, which manage to exploit many diverse habitats, one species (Cryptoblepharis boutonii) is found in the zone of exposed rocks near the ocean. This is a relatively harsh environment, containing virtually no vegetation. Another lizard family, the Geckonidae, has also been fairly successful in the American Samoas. The most common gecko is Gehyra oceanica, which attains the exceptional length of twelve inches. The four other species that were encountered are: Hemidactylus frenatus, Peropus mutilatus, Lepidodactylus lugubris, and, Cyrtodactylus pelagicus. One persistent pest that Kelly encountered while camping on the beach was the hermit crab. These crabs ate through plastic bags in an effort to devour food and herpetological specimens. I would like to thank Kelly for his informative presentation concerning the flora and fauna of beautiful American Samoa.

Next, Joseph T. Collins showed us some slides of reptiles that are not commonly encountered in many zoos. There was one slide of a young Loggerhead sea turtle (Caretta caretta) that was being held temporarily at the Cincinnati Zoo after it was confiscated from an individual that obtained it while vacationing in Florida. One remarkable slide depicted a South American worm lizard (Amphisbaena alba). Collins was able to obtain a good slide of a Black Mamba (Dendroaspis polylepis) that was nine feet in length. We also had the opportunity to see a rare python, the Black-headed Rock Snake (Aspidites melanocephalus), which was born in captivity at the Dallas Zoo. The slide show ended with a beautiful picture of a Green Tree Python (Chondrophython viridis) that had a blue dorsal stripe.

The meeting ended after Dr. Robert Clarke showed his slides. Among the more remarkable scenes, was that of a Cottonmouth (Agkistrodon piscivorus) swimming in the Dismal Swamp. Unlike the harmless watersnakes (Natricinae), the Cottonmouth holds much of its body on the surface or actually out of the water. There were also slides of Black Rat Snakes (Elaphe obsoleta) emerging from eggs, and, a Hognose snake (Heterodon sp.) regurgitating a toad (Bufo sp.).

---HANK GUARISCO, Museum of Natural History, University of Kansas, Lawrence, KS



AN UNUSUAL STRAND IN THE ANURAN FOOD-WEB

Last summer I acquired a juvenile African Bullfrog (Pyxicephalus adspersus) and soon became intrigued with its voracious appetite, which could be satisfied by supplying a wide variety of prey items of appropriate size. The frog, which is presently 4½ inches in length, can consume an adult mouse. When it becomes mature it could possibly reach a length of 9 inches and devour a young rat. Turning to the literature, I discovered that others have also been impressed by this frog's catholic tastes. One specimen, which is housed in the Transvaal Museum, was preserved in the act of eating a small chicken. Another, from the same collection, had swallowed seventeen young Ringhals cobras (Hemachatus haemachatus)! This is a truly remarkable feat, especially after seeing a picture of the frog and the snakes that were taken from its stomach. Further inquiry into the habits of the African Bullfrog revealed that recently metamorphosed individuals will overpower and consume one another, especially under crowded conditions.

Branch, W. R.

1976. "Two Exceptional Food Records for the African Bullfrog, Pyxicephalus adspersus (Amphibia, Anura, Ranidae)." J. Herpetol. 10(3):266-268.

Wager, V. A.

1965. The Frogs of South America. Purnell & Sons (S.A.) Pty., Ltd., Capetown, p. 130-134.

---HANK GUARISCO, Museum of Natural History, University of Kansas, Lawrence, KS.

* * * * *

COLLINS PHOTOS IN NEW AUDUBON BOOK

Joseph T. Collins, vertebrate zoologist at the KU Museum of Natural History in Lawrence and long-time KHS member, has 65 of his color photographs reproduced in the new Audubon Society "Field Guide to North American reptiles and amphibians," written by John Behler and Wayne King. The book was published by Alfred A. Knopf (New York) and is available at most book stores for \$9.95.

Many KHS members will find this book useful and informative. A number of Collins' photographs are of Kansas reptiles, and some were collected by KHS members. These are listed below.

Photo #326 Alligator Snapping Turtle -- This specimen is currently on exhibit at the Topeka Zoo.

Photo #366 Northern Earless Lizard -- Collected by James L. Knight and J. T. Collins in Cheyenne County, Kansas.

Photo #375 Northern Prairie Lizard -- Collected by Stanley Roth in Morton County, Kansas.

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- Photo #423 Northern Prairie Skink -- Collected by J. T. Collins in Jefferson County, Kansas.
- Photo #428 Southern Prairie Skink -- Collected by Eric Rundquist in Barber County, Kansas.
- Photo #429 Southern Coal Skink -- Collected by Mary Dawson and J. T. Collins in Douglas County, Kansas.
- Photo #455 Western Slender Glass Lizard -- Collected by J. T. Collins in Douglas County, Kansas.
- Photo #460 Plains Blackhead Snake -- Collected by Eric Rundquist and Kelly Irwin in Comanche County, Kansas.
- Photo #463 Flathead Snake -- Collected by Bruce, Kay and Mary Dawson in Wabaunsee County, Kansas.
- Photo #464 New Mexico Blind Snake -- Collected by Stanley Roth in Clark County, Kansas.
- Photo #480 Eastern Yellowbelly Racer -- Collected by James Knight and J. T. Collins in Cheyenne County, Kansas.
- Photo #497 Prairie Ringneck Snake -- Collected by J. T. Collins and Mary Dawson in Clark County, Kansas.
- Photo #506 Northern Redbelly Snake -- Collected by Ray Ashton, George Pisani and J. T. Collins in Jefferson County, Kansas.
- Photo #519 Graham's Crayfish Snake -- Collected by J. T. Collins in Douglas County, Kansas.
- Photo #534 Western Plains Garter Snake -- Collected by J. T. Collins in Ellsworth County, Kansas.
- Photo #543 Western Plains Garter Snake (red phase) -- Collected by Kelly Irwin, Marty Capron and Larry Miller in Barton County, Kansas.
- Photo #572 Plains Hognose Snake -- Collected by Mary Dawson and J. T. Collins in Comanche County, Kansas.
- Photo #577 Kansas Glossy Snake -- Collected by Stanley Roth and Ray Ashton in Morton County, Kansas.
- Photo #600 Central Plains Milk Snake -- Collected by Eric M. Rundquist in Comanche County, Kansas.
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Photo #615 Red Milk Snake -- Collected by Mary Dawson in Douglas County, Kansas.

Photo #638 Western Massasauga -- Collected by J. T. Collins and Mary Dawson in Comanche County, Kansas.

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MAMMALIAN HORMONE INDUCES EGG DEPOSITION IN TURTLES

Several years ago, it was discovered that oxytocin, a hormone secreted by the posterior pituitary gland (neurohypophysis) of mammals, induced egg deposition in turtles of many different species. In mammals, this hormone stimulates uterine contractions, especially toward the end of gestation. Arginine vasotocin and mesotocin, two reptilian counterparts of oxytocin, presumably aid the process of giving birth or laying eggs. It is known that captive turtles may sometimes retain their eggs for an excessively long period, during which time many of them spoil. Therefore, oxytocin can be a useful tool in preventing this condition. It may also be used to obtain eggs from many individuals at approximately the same time, which can be especially useful in research.

Basically, the procedure consists of the following steps: 1) injecting between .2 units and 10 units of oxytocin per 100 grams of turtle body weight. Lower doses are ineffective in inducing egg laying, while higher doses may actually delay egg deposition by causing the eggs to bunch up near the cloaca. 2) Within two to eight hours egg deposition is usually completed. Before injecting the hormone, however, the experimenter should be relatively certain that the eggs have been completely developed, i.e., that they have complete shells. This can be done by feeling the eggs through the wall of the inguinal pocket (the junction between the hind leg and the body).

Oxytocin has successfully induced egg deposition in turtles belonging to the following families: Kinosternidae, Chelidae, Pelomedusidae, Testudinidae, Chelydridae, Emydidae, and, Trionychidae. Furthermore, most of the eggs obtained by this method have successfully hatched.

Ewert, M. A. and J. M. Legler.

1978. "Hormonal Induction of Oviposition in Turtles." Herpetologica 34(3):314-318.

---HANK GUARISCO, Museum of Natural History, University of Kansas, Lawrence, KS.

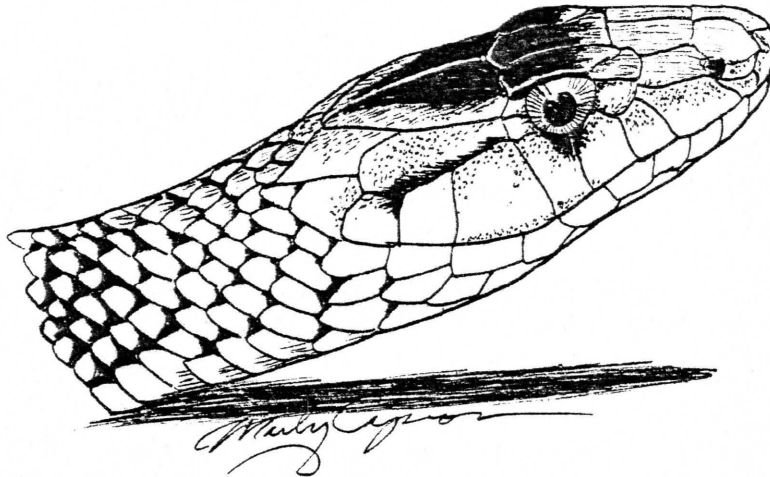
18 FOOT-LONG LIZARD FOUND IN NEW GUINEA

LONDON (AP) - British explorers say they have found the world's biggest lizard, an 18 foot-long reptile living in a swamp in Papua, New Guinea.

"It has large teeth and claws and local people call the lizard a dragon and say it eats people." explorer John Blashford-Snell said at a news conference.

He said members of the expedition that found the lizard are "still out there, observing it, so I expect some more details later."

A reptile expert at the British Natural History Museum said it could be a member of a newly discovered species. The Guinness Book of Records says there have been reports of New Guinea lizards measuring 15 feet 7 inches long, but that the longest lizard measured by scientists was 10 feet 2 inches long.



CARE IN CAPTIVITY SHEET NO. 7 -- CHICAGO HERPETOLOGICAL SOCIETY

CARE AND HATCHING OF REPTILE EGGS

INTRODUCTION -- Hatching of reptile eggs offers a rewarding challenge, and an opportunity to record important data about egg clutch numbers, egg sizes, egg growth, incubation times, hatching sizes, coloration differences from adults, etc. Freshly killed D.O.R. specimens sometimes offer a good source of eggs and an opportunity to hatch reptiles which would certainly be doomed otherwise.

SOURCES -- Suspected gravid captive reptiles require some additional care in preparation for the day the eggs will be laid. The expectant female will very often refuse to feed well. She may become very nervous, irritable and may be more prone to bite. I have observed on several occasions even the usually gentle smooth green snake become aggressive while carrying eggs. Provide a secure hiding place and avoid all unnecessary handling of the expectant female. Check her cage at least twice daily so that any eggs laid can be retrieved before they dry out (which would be fatal to the embryo). Turtle and terrapin eggs that are laid in the water must be retrieved immediately or the embryo will drown.

REQUIREMENTS -- Presuming that the eggs you have are fertile, the only other requirements for hatching the eggs successfully are:

1. A hatching medium in which to place the eggs that is sterile and holds moisture well.
2. An area in which to keep the eggs at a temperature between 70 and 90°F.
3. As little handling of the eggs as necessary during incubation.

FERTILITY AND GROWTH -- Determining if the eggs you have are fertile is usually not possible until the eggs have been in the hatching medium for 10 to 14 days. After this time, you may examine the eggs in a darkened room by shining the light of a small penlight through the eggs. Even though you may not detect signs of growth within the eggs yet, chances are good the eggs may be fertile and developing if there are no signs of spoilage. Infertile eggs will usually discolor, become hard and emit a foul odor within two weeks.

HATCHING MEDIA -- The following hatching mediums have been used with excellent success.

1. Perlite -- A granular pure white material much like vermiculite and obtainable very cheaply at large garden shops.
2. Vermiculite
3. Cotton
4. Sphagnum Moss
5. Paper Towels.-- Work well but tend to cause mold easily.

PROCEDURES -- I have had nearly 100% hatching success using Perlite as a hatching medium and recommend its use. You will need a container for the hatching medium. A styrofoam box, clay flower pot or a plastic bottle which can easily be cut to a height which will allow at least two inches of perlite on all sides of the eggs in the container. Place the perlite in the container first and add enough water to make the perlite damp (not wet). If the perlite sticks to the sides of the container, it's too wet. The perlite should fill the container to within one or two inches of the top. Bury the eggs in the perlite making sure there is at least two inches on all sides of the eggs. Place a lid on the container which allows air to circulate within the container. A few sheets of newspaper laid loosely on top of the container work fine. Watch the eggs closely for about the first three weeks. If mold occurs on any of the eggs, there's probably too much moisture. This mold can be removed with a cotton swab dipped in a mild antiseptic such as Listerine. Too much moisture will also cause the eggs to swell, eventually splitting them open, possibly killing the embryo. Eggs that are growing well should appear full or with a small depression or dent somewhere on each egg. If the eggs have numerous depressions or appear wrinkled, they're too dry. It is better to have too little than too much moisture. Moisture which is lost through evaporation must be replaced periodically. This moisture is best added by spraying the top of the perlite with a fine spray of water. Plastic bottles with adjustable spray heads are available in garden shops and work excellently.

TEMPERATURE -- Most eggs should be kept at a room temperature between 70 and 90°F. A room temperature that rises during the day between 80 and 90°F with a nighttime cooling is preferred to a constant higher temperature. Never subject the eggs to a temperature over 95°F for more than brief periods.

Consider the species' natural range when determining the average temperature in which you will keep the incubating eggs. Reptile eggs from species found in warmer climates should be kept at the higher end of the suggested temperature. Eggs from species found in arid regions usually require slightly less moisture during incubation.

DEVELOPMENT DURATION -- Most reptile eggs will hatch within 45 to 75 days, although some species will hatch after a much shorter incubation. Whenever possible it is best to release most of the hatchlings in their native habitat.

AUTHOR FOR THIS ACCOUNT -- TOM ZAREMBA

THIS CARE IN CAPTIVITY SHEET IS PUBLISHED BY THE CHICAGO HERPETOLOGICAL SOCIETY. PRIMARY EDITOR FOR THIS ACCOUNT IS STANLEY DYRKACZ.

The Chicago Herpetological Society is a non-profit organization incorporated under the laws of the state of Illinois. It's purposes are education, conservation and the advancement of herpetology. Meetings are held on the last Wednesday of each month (except November and December) at the Chicago Academy of Sciences, 2001 N. Clark St., Chicago, IL 60614. Care in Captivity sheets may be purchased in bulk quantity for about \$.05 each, by writing to the CHS, 2001 N. Clark St., Chicago, IL 60614.

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1976

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DO SOMETHING ABOUT SPRAY DRIFT

On May 30 the Friends of the Earth formally petitioned the Environmental Protection Agency and the Federal Aviation Administration for new pesticide labels and regulations. The purpose is to make it illegal to let pesticide drift upon people or their property without prior written permission. FOE is also asking the FAA to adopt a point system for determining when a pilot's license should be revoked for the damage he has caused with pesticides.

EPA and the FAA have just published a joint notice in the Federal Register (January 17) asking for public comment. The FAA would like comments in duplicate sent to: FAA Office of the Chief Counsel, Attn: Rules Docket (AGC-24), Docket No. 19448, 800 Independence Ave., SW, Washington, D.C. 20591. EPA would like comments (duplicates not necessary) sent to: Document Control Officer, Chemical Information Division (TS-793), FAA Docket No. 19448, Environmental Protection Agency, Room 447 East Tower, 401 M St., SW, Washington, D.C. 20460.

THE PETITION TO EPA

FOE is asking EPA to: 1) Adopt labelling regulations which would require something like the following to appear on all pesticide labels. (The instructions on pesticide labels have the force of law.)

WARNING

This pesticide must not be allowed to drift onto people or property without prior written permission by that person or property owner. For aerial application, at minimum written permission must be obtained from all persons living or owning property within 1,000 feet of the spray boundary. For ground rig operation other than fine droplet misting, the applicator must obtain at minimum written permission from all persons living or owning property within 250 feet of the spray project boundary -- 1,000 feet for misting.

2) Recognize that people have a right not to be sprayed with a pesticide without their permission. 3) Harmonize with the FAA program (as described below) and develop with the FAA and USDA a model applicator training program for all federally sponsored spray programs. 4) Forbid the sale of any pesticide so volatile that drift can never be controlled by any applicator. 5) Include integrated pest management material on all pesticide labels. (Integrated pest management is an ecologically-based strategy which places maximum reliance on host resistance and biological, cultural and mechanical control of pests with regular field monitoring. It minimizes the use of chemical pesticides.) For example:

INTEGRATED PEST MANAGEMENT

An integrated pest management program for your crop may be available from your local university or from the Extension Service of the USDA.

6) Create a Citizen Enforcement and Monitoring Package with Xerox cards and instructions for location to catch spray drift, and where to process them. 7) Create an appeals structure within EPA so citizens may get a second opinion from higher authority.

THE PETITION TO THE FAA

FOE's petition to the FAA asks them to: 1) Put someone in charge of protecting people from spray drift as required by the regulations which forbid "creating a hazard to persons or property on the surface." 2) Rather than exempting federal pilots from its regulations, develop a model applicator and monitoring program with EPA and the USDA for all federally sponsored spray programs. 3) Sharply adjust its present requirements relating to the size of identification numbers on aerial spray aircraft. 4) Adopt a point system for regulating spray that would encourage person to person agreements. Each person with an operator's license would lose his agricultural aircraft operator

certificate when he accumulates 10 current violation points. Points would be erased 3 years after they are assessed to the license. The following are examples of the penalties suggested by FOE:

For aerial spraying without first obtaining written permission of persons and property owners within 1,000 feet - - - 1 point

For causing a person to be hospitalized by allowing spray to drift on him, in the absence of previous written permission - - - 10 points

For spraying a schoolbus stop or community facility - - - 10 points

For spray of an automobile on the public highway - - - 5 points

COMMENTS

To be considered, comments must be submitted to the agencies on or before April 17, 1980. The two agencies are inviting all comments, but would especially like to see the following issues addressed: 1) The appropriate role for the FAA under a comprehensive Federal program of agricultural aircraft operation, including any specific responsibilities in the aerial application of pesticides. 2) Any changes or additions to the FAA rules governing the flight safety of agricultural aircraft operations that are consistent with FAA statutory authority to regulate aeronautical activities. 3) The utility of placing more explicit application practices and use restrictions, e.g., nozzle sizes, buffer zones, in terms of target pests and sites, on pesticide labeling. 4) Is labeling an effective means of communicating information on integrated pest management and requirements for public notification? What other methods would be appropriate? 5) What mechanism could be employed to provide the public with advance notice of broadscale spray programs? Should requirements for advance notice of spraying and obtaining written permission to spray from adjoining property owners be applied equally to broadscale spray programs and local agricultural applications?

Certainly, any personal experiences with spraying and the FAA's non-regulation of aerial spraying would be very effective. But there are reasons for everyone to be concerned: spray which is not confined to the target area may kill or injure bees, livestock, people, trees, or other plants; it may disrupt other methods of pest control; and it may contaminate air and water.

For further information, see the January 17 Federal Register. I would be happy to tell anyone who is interested of our experience with the FAA. Do you have other ideas for protecting people and property from spray drift? Please let me know.

---TERRY SHAFER, R.R. 3, Lawrence, KS 66044. Phone: 913/842-1348

KANSAS SIZE RECORD FOR BLACK RAT SNAKE

While advising an Avila College herpetology class field trip on 24 April 1977, Harry Gregory captured a record length black rat snake (Elaphe obsoleta obsoleta) at Butler County State Park, Butler County, Kansas. It was found along the stream bed below the spillway. The male snake measured 2054 mm (80.9 inches) when alive, and 1880 mm (74 inches) after preservation. The specimen, deposited in the Bobby Witcher Memorial Collection at Avila College, Kansas City, Missouri (BWMC 00551), has been donated to the Museum of Natural History, University of Kansas, Lawrence.

---ROBERT POWELL, Department of Natural Sciences, Avila College, Kansas City, Missouri, 64145

REGIONAL HERPETOLOGICAL SOCIETIES

In the interest of furthering the cooperation among herp fanciers, the following list of regional herpetological societies is provided.

ARIZONA HERP. ASSOC.
Grace Harris, Secretary
4028 North 55th Ave.
Phoenix, Arizona 85031

CONNECTICUT HERP SOCIETY
Donna Douglas
790 Orchard St.
New Haven, Connecticut 06511

ARKANSAS HERP. SOCIETY
5312 West 13th St.
Little Rock, Arkansas 72205

DALLAS HERP SOCIETY
Wayne Seifert
Dallas Museum of Natural History
P.O. Box 26193
Dallas, Texas 75226

CANADIAN AMPHIBIAN & REPTILE
CONS. SOCIETY
Barbara Froom
8 Preston Place
Toronto, Ont., Canada M4N 2S9

EASTERN SEABORD HERP LEAGUE
Malvin Skaroff
1025 Lakeside Avenue
Philadelphia, Pennsylvania 19126

CENTRAL OHIO HERP SOCIETY
887 Oxley Road
Columbus, Ohio 43212

FLORIDA HERP SOCIETY
Dennis R. Magee
5488 North River Road
Jacksonville, Florida 32211

CHICAGO HERP SOCIETY
c/o Chicago Academy of Sciences
2001 North Clark St.
Chicago, Illinois 60614

FLORIDA WEST COAST HERP SOCIETY
John Lewis
1312 South Evergreen Ave.
Clearwater, Florida 33515

COLORADO HERP SOCIETY
P.O. Box 15381
Denver, Colorado 80215

GEORGIA HERP SOCIETY
c/o Reptile House
Atlanta Zoological Park
800 Cherokee Ave. S.E.
Atlanta, Georgia 30315

GREAT LAKES HERP SOCIETY

James Todd
15554 Cooper
Taylor, Michigan 48180

GREATER CINCINNATI HERP SOCIETY
Cincinnati Museum of Natural History
172 Gilbert Avenue
Cincinnati, Ohio 45202

GULF COAST HERP SOCIETY

John Zapata
P.O. Box 1562
Houston Zoo
Houston, Texas 77001

IOWA HERP SOCIETY

P.O. Box 191
Norwalk, Iowa 50211

KANSAS HERP SOCIETY

Hank Gaurisco
Museum of Natural History
University of Kansas
Lawrence, Kansas 66045

KENTUCKY HERP SOCIETY

John MacGregor
102 Fourth St.
Nicholasville, Kentucky 40356

MARYLAND HERP SOCIETY

Natural History Soc. of Maryland
2643 North Charles St.
Baltimore, Maryland 21218

MASSACHUSETTS HERP SOCIETY

P.O. Box 1082
Boston, Massachusetts 02103

MID-MISSISSIPPI VALLEY HERP SOC.

Mike Lodato
925 Park Plaza Drive
Evansville, Indiana 47715

NEBRASKA HERP SOCIETY

2326 S. 12th St.
Omaha, Nebraska 68108

NEW MEXICO HERP SOCIETY

Department of Biology
University of New Mexico
Albuquerque, New Mexico 87131

NEW YORK HERP SOCIETY

David Hulmes
122 Second Avenue
Hawthorne, New York 07507

NORTH CAROLINA HERP SOCIETY

Patricia Ashton
N.C. State Museum of Natural History
101 Halifax St.
Raleigh, North Carolina 27611

NORTH NEW JERSEY HERP SOCIETY

P.O. Box 94
Augusta, New Jersey 07822

NORTHERN OHIO ASSOC. HERPS

Department of Biology
Case Western Research University
Cleveland, Ohio 44106

OKLAHOMA HERP. SOCIETY

Jeffrey Black
Department of Biology
Oklahoma Baptist University
Shawnee, Oklahoma 74801

PACIFIC NORTHWEST HERP. SOC.

Frank Slavens
P.O. Box 30744
Seattle, Washington 98103

PALM BEACH COUNTY HERP. SOC.

Greg Longhurst
P.O. Box 125
Loxahatchee, Florida 33470

PHILADELPHIA HERP SOCIETY

John Robinson
ANSP, 18th and the Parkway
Philadelphia, Pennsylvania 19126

SACRAMENTO VALLEY HERP. SOC.

Bob Pedder
6007 Watt Avenue
North Highlands, California 95660

ST. LOUIS HERP. SOCIETY
P.O. Box 9216
St. Louis, Missouri 63117

SOUTH FLORIDA HERP. SOC.
Saul Friess
9972 N. Kendall Drive #54
Miami, Florida 33176

SOUTHERN ARIZONA HERP. SOC.
Tom Boyden
4521 W. Mars St.
Tucson, Arizona 85704

SOUTHWESTERN HERP. SOCIETY
San Fernando Valley Chapter
Rick Korne
5730 Newcastle Avenue
Encino, California 91316

SOUTHWESTERN HERP. SOCIETY
Bob Sanders
San Bernadino County Museum
2024 Orange Tree Lane
Redlands, California 92373

TEXAS HERP. SOCIETY
Barry Hinderstein
8739 Link Terrace
Houston, Texas 77025

UTAH HERP. SOCIETY
Hogle Zoological Gardens
P.O. Box 8475
Salt Lake City, Utah 84108

VIRGINIA HERP. SOCIETY
Franklin J. Tobey, Jr.
P.O. Box 1376
Leesburg, Virginia 20075

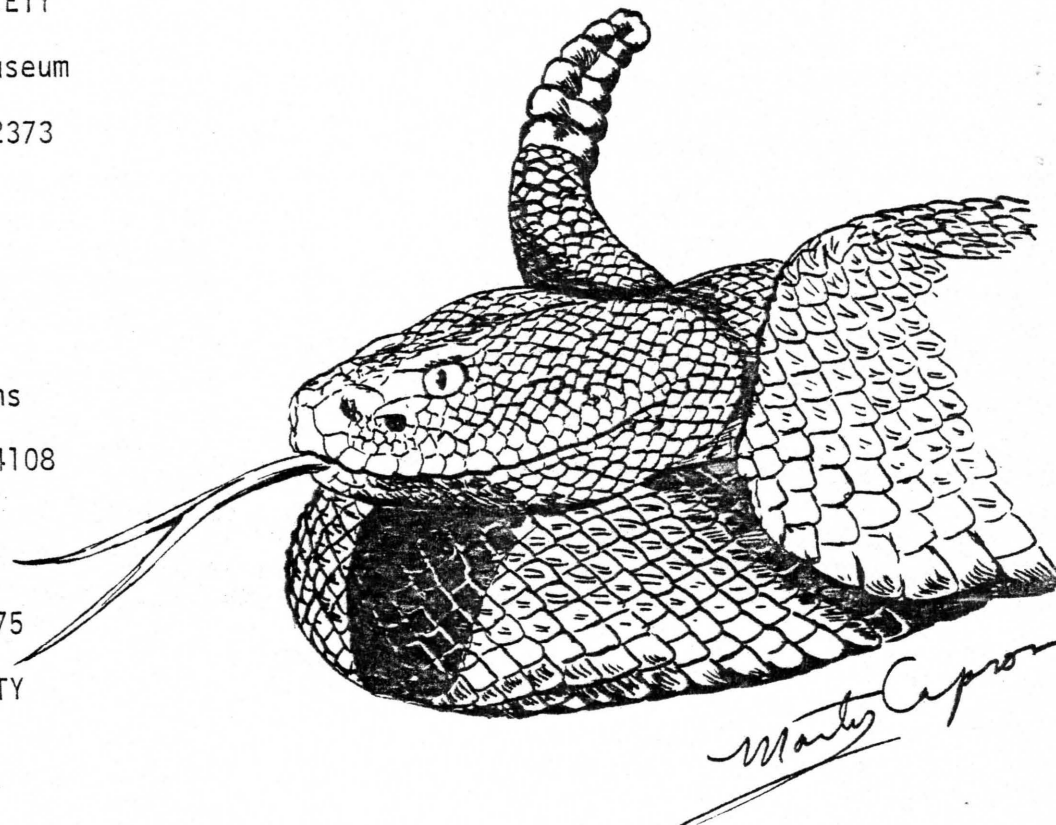
WASHINGTON HERP. SOCIETY
Scott Rae
317 Adahi Road, S.E.
Vienna, Virginia 21180

WESTERN MASS. HERP. SOC.
c/o Science Museum
236 State St.
Springfield, Massachusetts 01103

WISCONSIN HERP. SOCIETY
Milwaukee Public Museum
800 West Wells St.
Milwaukee, Wisconsin 53233

Robert Wagner
AAZPA - Oglebay Park
Wheeling, West Virginia 26003

SAN DIEGO HERP SOCIETY
Don Hunsaker
Dept. of Zoology
San Diego State University
San Diego, California 92115



A NOTE TO MEMBERS

Please take a few minutes to check the accuracy of your name and address in the following membership list. It is very important that we have your complete and accurate address. Bulk mail is not forwardable, so you must notify us immediately if you have a change of address.

We have had a few problems with the mail service during the past year. You should receive your newsletter about the 20th of each following a meeting. Please notify the KHS Editor or me if you do not receive future newsletters.

---LARRY MILLER, Secretary-Treasurer, 524 North Osage Street, Caldwell, KS 67022

REGULAR MEMBERS

Kraig Adler, Langmuir Laboratory, Cornell University, Ithaca, NY 14853
 Brad Anderson, 420 West Broadway, Newton, KS 67114
 Larry Andrews, 318 South Pine, Ponca City, OK 74601
 John Applegarth, Dept. Biology, Univ. New Mexico, Albuquerque, NM 87131
 John Atherton, 1813 N. St. Paul, Wichita, KS 67203
 Bob Bader, Route 2, Box 78, Brookneal, VA 24528
 Marja van Barneveld, Taalstr 31, 5261 BA Vught, NETHERLANDS
 George Berline, Route 2, Box 77, Caldwell, KS 67022
 Warren Bird, 2020 Thompson, Dodge City, KS 67801
 Jeffrey H. Black, Dept. Biology, Oklahoma Baptist Univ., Shawnee, OK 74801
 Walter Boles, 2007 Briarcliff Lane, Emporia, KS 66801
 Randy Boudeman, 3825 S.E. 31st, Topeka, KS 66605
 Jens Petter Brastad, Brusetskollen 16, 1364 Hualstad, NORWAY
 Jeffrey T. Burkhart, Dept. Biology, St. Mary of th Plains Coll., Dodge City, KS 67801
 Melissa Burt, West 828 25th St., Spokane, WA 99203
 Louis J. Bussjaeger, 2001 S. Gold, Wichita, KS 67213
 Edward R. Byrne, c/o Weed Park Zoo, City Hall Build., Dept. of Parks,
 & Recreation, Muscatine, IA 52761
 Jan Caldwell, Savannah River Ecology Lab., Drawer E, Aiken, SC 29801
 David Cannatella, Museum of Natural History, Univ. Kansas, Lawrence, KS 66045
 Martin B. Capron, Box 542, Oxford, KS 67119
 Jane Carruthers, 2432 N. Jackson, Wichita, KS 67204
 Park W. Carter, Box 145, Eureka, KS 67045
 Bob Clark, 9709 Sagamore Rd., Leawood, KS 66206
 Gary K. Clarke, Topeka Zoo, 635 Gage Blvd., Topeka, KS 66606
 John W. Clarke, Clarke Environmental Services, 4801 W. Irving, Wichita, KS 67209
 Robert F. Clarke, Dept. Biology, Emporia State University, Emporia, KS 66801
 Mike Coker, Topeka Zoo, 635 Gage Blvd., Topeka, KS 66606
 Keith Coleman, 1916 Atwood, Topeka, KS 66604
 Barbara Coler, 822 Canterbury Lane, Lawrence, KS 66044
 Joseph T. Collins, Museum of Natural History, Univ. Kansas, Lawrence, KS 66045
 Dennis Cote, Route 2, Box 75, Clifton, KS 66937
 Adrian Crane, R.D. Box 235, Eatontown, NJ 07724
 Philippe A. Delepaul, 23, Roe de Mouscron, 7760 Dottignies, Belgium-Belgique

Patricia Devine, 9211 Nieman Rd., Overland Park, KS 66214
Jeff DeWeese, P.O. Box 153, Wellington, KS 67152
Dewayne Dick, P.O. Box 3223, Bartlesville, OK 74003
Conne Dillon, Sedgwick County Zoo, 5555 Zoo Boulevard, Wichita, KS 67212
Michael L. Dohanec, 19 Smedley St., North East, PA 16428
William E. Duellman, Museum of Natural History, Univ. Kansas, Lawrence, KS 66045
Dean Dwigans, 617 West 59th Terr., Kansas City, MO 64113
Stanley Dyrkacz, 307 South Hickory, McPherson, KS 67460
Jeff Ehlers, 218 N. St. George St., Caldwell, KS 67022
Victor B. Eichler, Dept. of Biology, Box 26, Wichita State Univ., Wichita, KS 67208
William K. Engelder, 153 Scott Ave., Wellsville, NY 14895
Bobbie Espinosa, 817 S. Fountain, Wichita, KS 67218
James D. Fawcett, Dept. of Biology, Univ. Nebraska-Omaha, Box 688, Omaha, NE 68182
Andrew Finfrock, 18680 Faure Ridge Rd., Los Gratos, CA 95030
Henry S. Fitch, Rt. 3, Box 142, Lawrence, KS 66044
Virginia R. Fitch, Rt. 3, Box 142, Lawrence, KS 66044
Thomas L. Flowers, 803 Morrison, Dodge City, KS 67218
Jim Foral, 2718 Everett, Lincoln, NE 68502
John C. Fraser, 119 North 15th St., Fredonia, KS 66736
Arnold Froese, Dept. Ind. & Group Behavior, Sterling Coll., Sterling, KS 67579
Darrel Frost, Museum of Natural History, Univ. Kansas, Lawrence, KS 66045
Greg M. Goff, Route 5, Box 34, Lawrence, KS 66044
Ronald Gonzales, 921 E. Kaweah, Visalia, CA 93277
Thomas A. Graham, 920 N. Summit, Arkansas City, KS 67005
Peter Gray, Museum of Natural History, Univ. Kansas, Lawrence, KS 66045
Harry L. Gregory, Kansas City Zoo, Swope Park, Kansas City, MO 64132
David Grow, 9508 N. Walker, Apt. 230N, Oklahoma City, OK 73114
Terry B. Grund, 941 Pennsylvania, Lawrence, KS 66044
Hank Guarisco, P.O. Box 3171, Lawrence, KS 66044
Hanne Guarisco, Gartenstr. 25, D5410 Hohn - Grenzhausen, WEST GERMANY
Richard Hager, 314 E. Laura, Salina, KS 67401
Kelly Haller, 17909 West 79th St., Lenexa, KS 66219
Jeff Harshfield, 1539 S. Pattie, St., Wichita, KS 67211
Harold E. Hedges, 4501 Francis St., Kansas City, KS 66103
Roger A. Henry, R.R. #1, Box 192, Smithshire, IL 61478
Aaron Henton, Box 154, South Haven, KS 67140
Gus Huey, 109 N. Broadmoor, Topeka, KS 66606
Thomas A. Huff, Reptile Breeding Foundation, P.O. Box 1450, Picton,
Ontario KOK 2T0 CANADA
James Irwin, 2218 West 2nd St., Topeka, KS 66606
Kelly Irwin, 2218 West 2nd St., Topeka, KS 66606
James D. Jennings, 1711 N. Madison St., Hutchinson, KS 67501
Jerry D. Johnson, Dept. of Biology, El Paso Community Coll., El Paso, TX 79998
Randall N. Johnson, 2805 West San Miguel, Phoenix, AZ 85817
Robert R. Johnson, 8528 Goodman Dr., Overland Park, KS 66212
Tom R. Johnson, Missouri Dept. of Conservation, P.O. Box 180, Jefferson City, MO 65101
Troy Johnson, 603 North Ewing, Caldwell, KS 67022
Al Kamb, 2619 Missouri St., Lawrence, KS 66044
Ray Keith, 3825 S. Seneca #84, Wichita, KS 67217
Brandon Keltner, 437 Westfield Ct., Wichita, KS 67212

Denny Kilgore, 2936 West St. Moritz, Phoenix, AZ 85023
James L. Knight, Museum of Natural History, Univ. Kansas, Lawrence, KS 66045
Dennis Kramer, Brewster, KS 67732
Rose Etta Kurtz, Museum of Natural History, Univ. Kansas, Lawrence, KS 66045
Janet M. Labrecque, 5307 West 99th Terr., Shawnee Mission, KS 66207
Jim Lam, 5130 N.W. Pueblo, Topeka, KS 66606
Richard L. Lardie, 313 Flintridge Rd., Enid, OK 73701
Richard Lattis, Bronx Zoo, 185th & Southern Blvd., Bronx, NY 10460
Juan R. Leon, Departamento de Biología, Universidad de Oriente, Cumana,
Edo. Sucre, VENEZUELA, S.A.
Brian Levenson, 877 Roosevelt St., West Hempstead, NY 11552
Stephanie F. Lyall, P.O. Box 150, Eudora, KS 66025
Larry & Angie Madl, 1517 N. Heorman, Andover, KS 67002
Dennis R. Magee, 5488 N. River Rd., Jacksonville, FL 32211
Luis Malaret, Museum of Natural History, Univ. Kansas, Lawrence, KS 66045
Jim Marlett, Sedgwick County Zoo, 5555 Zoo Blvd., Wichita, KS 67212
Danny McClung, 1310 South 39th Place, #55, Muskogee, OK 74401
Delfi Messinger, 9900 David Moore Drive, Austin, TX 78748
Clyde G. Miller, 617 Morningside, Wellington, KS 67152
Frank Miller, 7303 Church St., Morton Grove, IL 60053
Larry Miller, 524 North Osage Street, Caldwell, KS 67022
Michael J. Miller, D.V.M., 5223 S. Francisco, Chicago, IL 60632
Zeda Miller, 617 Morningside, Wellington, KS 67152
Kirk Mullen, 6227 Eilerts, Wichita, KS 67218
Eric Murray, 3810 S.E. 32nd Place, Topeka, KS 66605
Pierre Musy, 9409 West 97th Terr., Overland Park, KS 66212
Marc Nadeau, Museum of Natural History, Univ. Kansas, Lawrence, KS 66045
Kyle Newkirk, 22 North Young Street, Box 249, Caldwell, KS 67022
Scott Palmer, 1805 Cherryhurst, Houston, TX 77006
Barbara Paschke, 731 Tennessee, Lawrence, KS 66044
Michael R. Patterson, 8 Royal Drive, Kansas, KS 66111
Janice Perry, Dallas Zoo, 621 East Clarendon Dr., Dallas, TX 76203
John Perry, 16455 S.W. Myrtle Avenue, Tigarde, OR 97223
Marjorie Perry, Ace Ranch #10, 4453 San Antonio Hwy., Kerrville, TX 78028
Jim Pilch, 808 West 27th, Lawrence, KS 66044
George Pisani, Biology Dept., Univ. Kansas, Lawrence, KS 66045
Dwight R. Platt, R.F.D. #2, Box 209, Newton, KS 67114
Michael V. Plummer, Dept. of Biology, Harding College, Searcy, AR 72143
Ron Polett, Rt., #1, Box 132, Coffeyville, KS 67337
Robt. Powell, 2725 28th St. Court, Independence, MO 64055
Gregory Pregill, Div. Reptiles & Amphibians, Smithsonian Institution, Washington,
D.C. 20560
Scott Ratliff, 2206 West MacArthur, #131, Wichita, KS 67217
Stan Roth, 532 Oklahoma St., Lawrence, KS 66044
Gareth Rowell, 450 Nebraska St., Lawrence, KS 66044
John A. Ruiz, 1624 Lomax Lane, Redondo Beach, CA 90278
Eric M. Rundquist, Oklahoma City Zoo, Oklahoma City, OK 73114
Allen H. Savitzky, Ecology & Systematics, Langmuir Lab, Cornell Univ., Ithaca,
NY 14853
Terry D. Schwaner, 195 Cindy Lou Place, Mandeville, LA 70448
Frank Powell, 1928 Idlewild, Wichita, KS 67216

Marvin D. Schwilling, Game Research Office, 1803 West 6th, Emporia, KS 66801
John Simmons, Dept. of Herp., California Acad. Sciences, Golden Gate Park,
San Francisco, CA 94118
Shelley Skie, 2815 Iowa, Lawrence, KS 66044
Greg Skrdla, R.R. #2, Box 50, Caldwell, KS 67022
Hobart M. Smith, Dept. E.P.O. Biology, Univ. Colorado, Boulder, CO 80309
Allan W. Smits, 821 22nd Terr., Lawrence, KS 66044
Anthony Sokol, 7 Roma St., Bentleigh 3204, Victoria, AUSTRALIA
Wally L. Speers, 8518 Standing Rock, San Antonio, TX 78242
Joe Spurgeon, 4844 W. Ridge Rd., Wichita, KS 67205
Mark A. Stahle, 1000 4th St., New Cumberland, PA 17070
Chris Stammler, 914 West 27th Terr., Lawrence, KS 66044
Ned Stichman, 725 Washington St., Independence, KS 67301
Kemper Straley, 1810 Booth, Winfield, KS 67156
Richard A. Sugerman, 2308 N. Bluff, Wichita, KS 67220
J. R. Sutton, 1316 East 21st St., Lawrence, KS 66044
Shelley Tarbet, Okla. City Zoo, Rt. 1, Box 1, Oklahoma City, OK 73111
Max C. Thompson, Dept. Biology, Southwestern College, Winfield, KS 67156
George F. Toland, 908 Highland, Salina, KS 67401
John Tollefson, 2819 Main Ct., Lawrence, KS 66044
Gene Trott, 1116 East Lincoln St., Wellington, KS 67152
Jody Trott, R.R. #2, Box 58, South Haven, KS 67140
Toby Trott, 1116 East Lincoln, St., Wellington, KS 67152
Linda Trueb, Museum of Natural History, Univ. Kansas, Lawrence, KS 66045
Thomas G. Vermersch, 5118 Anna Maria, San Antonio, TX 78214
Joachim Walther, 1614 Ellis, Wichita, KS 67211
Kurt Walther, 1614 Ellis, Wichita, KS 67211
Robert Waltner, 2216 East 51st South, Wichita, KS 67216
Mark Warner, 1942 Judson St., Manhattan, KS 66502
Michelle Warner, 209 North Webb St., Caldwell, KS 67022
Rob Wencel, R.R. #3, Caldwell, KS 67022
Craig White, R.R. #2, Caldwell, KS 67022
Bernard Willard, 1217 Kentucky St., Lawrence, KS 66044
Tamara F. Williams, 1325 Massachusetts, Lawrence, KS 66044
Dennis W. Wilson, Bonner Springs High School, Box 216, Bonner Springs, KS 66012
Richard D. Worthington, Dept. Biol. Sci., Univ. of Texas, El Paso, TX 79968
Zoo Odyssey, 4276 Ft. Simmons Avenue, LaBelle, FL 33935

1979 CONTRIBUTING MEMBERS

Mary Palmer, 1805 Cherryhurst, Houston, TX 77006
W.E. de Ruyter, Margreitstr 11, 4571 US Axel, NETHERLANDS
Darrel Simmons, 3110 Kingston, Ponca City, OK 74601

NEW 1980 MEMBERS (as of 22 January 1980)

Martin Allen, 5330 North Wichita St., Wichita, KS 67204
 I.R. Atkinson, East Central St., Caldwell, KS 67022
 Jacquelin L. Barnes, 2032 Southwest Expressway #15, San Jose, CA 95126
 Mike Clausen, Box 215, South Haven, KS 67140
 Robert J. Gress, 3228 N. Oliver, Wichita, KS 67220
 Gean Ann Harris, 3906 S.E. 32nd. Topeka, KS 66605
 Carl Madorin, 139 N. Kendall, Topeka, KS 66606
 Ron Nagle, 4912½ N. Hayter Ave., Lakewood, CA 90712
 Patricia A. Neeland, Caldwell High School, USD #360, Caldwell, KS 67022
 Mathew C. Nowak, 1007 N. 2nd, Lansing, KS 66043
 Michael S. Rush, Dept. Biol. Sciences, Fort Hays State Univ., Hays, KS 67601
 Rich Siegel, Museum of Natural History, Univ. Kansas, Lawrence, KS 66045
 Six Grade Class, Caldwell Elementary School, USD #360, Caldwell, KS 67022
 Andrea Stammer, 914 West 27th Terr., Lawrence, KS 66044
 Ed Stegal, Sedgwick County Zoo, 5555 Zoo Blvd., Wichita, KS 67212
 Sara Swartz, 1501 El Camino, Ponca City, OK 74601
 Leland Unruh, 316 Western, Haysville, KS 67060
 Georgia I. Flauding, P.O. Box 72, South Haven, KS 67140
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NOW AVAILABLE!!

"Amphibians and Reptiles of the Carolinas and Virginia", by Bernard S. Martof, William M. Palmer, Joseph R. Bailey, and Julian R. Harrison III. Photography by Jack Dermid. 272 pp., \$14.95.

The only illustrated guide to the amphibians and reptiles of the Carolinas and Virginia. Here - in stunning color photographs and with up-to-date accounts - are the region's 159 species of frogs, snakes, turtles, salamanders, lizards, and alligator. Range maps indicate where each species is found.

A welcome guide and an important contribution to the natural history of the area, this book is essential for naturalists, backpackers, campers, hikers, and students of herpetology and general biology. And its a must for every library in our region.

(This publication available from the University of North Carolina Press, P.O. Box 2288, Chapel Hill, NC 27514)

SSAR GRANTS-IN-HERPETOLOGY

1980 SSAR Grants-in-Herpetology Deadline

The Society for the Study of Amphibians and Reptiles announces the continuation of its Grants-in-Herpetology program. Four grants of at least \$275 each will be available for awarding at the SSAR annual meeting in 1980 in Milwaukee, Wisconsin. Proposals will be considered in the following areas: 1) herpetology-oriented conservation, 2) graduate student research, 3) regional society programs or projects, and 4) research in zoos.

Proposals should be typewritten (no more than five double-spaced pages) and include a statement of the problem, objectives, methods, and budget. Each applicant should have a letter of support sent by a graduate advisor, institutional or organizational officer, or another person qualified to assess their proposal. Proposals will be evaluated on the basis of appropriateness to the program, chances of success, and qualifications of the applicant by a committee comprised of Linda R. Maxson, John D. Groves, Martin J. Rosenberg, Terry Graham, and Harry W. Greene. Proposals and supporting letters should be submitted in duplicate no later than 1 April 1980 to HARRY W. GREENE, Museum of Vertebrate Zoology, University of California, Berkeley, California 94720.

1979 RECIPIENTS OF SSAR GRANTS-IN-HERPETOLOGY

In 1978 the SSAR Board of Directors approved a Grants-in-Herpetology program. Its purpose is to financially support projects in four areas in which traditional sources of funding are not appropriate. Eight hundred dollars were budgeted for the first year, and an additional \$100 were received from the Northern Ohio Association of Herpetologists, permitting four grants of \$225 each.

Twenty-four proposals were received, including ten for graduate student research, three for zoo research, five for conservation, and six for regional society projects. These were carefully considered by a committee consisting of C. Kenneth Dodd, Martin J. Rosenberg, John Behler, Stephen R. Edwards, and Harry W. Greene. The following winners were announced at the annual meeting in Knoxville: Linda S. Ford (graduate student research, University of Texas at Arlington), Interspecific relationships among cloud forest Eleutherodactylus; John D. Groves (zoo research, Philadelphia Zoological Garden), The food and feeding behavior of the emerald tree boa, Corallus canina; Norman G. Reichenbach (conservation, Ohio State University), Ecology and management of the prairie garter snake, Thamnophis radix, in Ohio; and Wisconsin Herpetological Society (regional herp societies), Production of filmstrips for herpetological education.

---HARRY W. GREENE, Grants-in-Herpetology Committee Chairperson, Museum of Vertebrate Zoology, University of California, Berkeley, CA 94720.

(Taken from Herp Review 10(4), 1979)